

Acute kidney injury due to tropical infectious diseases and animal venoms – a tale of two continents

Emmanuel A. Burdmann MD, PhD <sup>1</sup>

Vivekanand Jha MD, DM, FRCP <sup>2</sup>

<sup>1</sup> LIM 12, Division of Nephrology, University of Sao Paulo Medical School, Sao Paulo, Brazil

<sup>2</sup> George Institute for Global Health, New Delhi, India and University of Oxford, Oxford, UK

Corresponding author: Emmanuel A. Burdmann, Faculdade de Medicina da Universidade de São Paulo, Av. Dr. Arnaldo, 455 – 3º Andar, sala 3310 (LIM 12), São Paulo, SP, Brazil , 01246-903. Phone: 55-11-30617343. Fax: 55-11-30882267. E-mail: [burdmann@usp.br](mailto:burdmann@usp.br)

EAB is partially supported by grants from “Conselho Nacional de Desenvolvimento Científico e Tecnológico - National Counsel of Technological and Scientific Development” (CNPq, 305858/2013-0) and “Fundação de Amparo à Pesquisa do Estado de São Paulo – São Paulo Research Foundation” (FAPESP, 2014/19286-4).

Running headline: Tropical diseases-associated acute kidney injury

## **Abstract**

South and Southeast Asia and Latin American together comprise 46 countries and are home to about 40% of the world population. The sociopolitical and economic heterogeneity, tropical climate and malady transitions characteristic of the region strongly influence disease behavior and healthcare delivery. AKI epidemiology mirrors these inequalities. In addition to hospital-acquired AKI in tertiary care centers, these countries face a large preventable burden of community-acquired AKI secondary to tropical infectious diseases or animal venoms, affecting previously healthy young individuals. This manuscript reviews the epidemiology, clinical picture, prevention, risk factors and pathophysiology of AKI associated with tropical diseases (malaria, dengue, leptospirosis, scrub typhus, and yellow fever) and animal venom (snakes, bees, caterpillars, spiders and scorpions) in tropical regions of Asia and Latin America, and discusses the potential future challenges due to emerging issues.

**Keywords:** acute kidney injury; Asia; Latin-America; tropical infectious diseases; animal venom; emerging issues.

## Introduction

The tropical regions of Asia (South and Southeast Asia), consist of 20 countries spread over 9.6 million km<sup>2</sup> and a population of 2.32 billion (33-58% urban). South and Southeast Asia have a GNI per capita (Atlas method) of about US\$1,496 and US\$ 9,602, respectively.<sup>1,2</sup>

Latin America (LA) comprises 26 countries in approximately 19.2 million km<sup>2</sup>, from Mexico and the Caribbean Islands in the north to Argentina and Chile in the south. Its population is estimated at 626 million (78% urban), with a GNI per capita of US\$9,095.<sup>3-6</sup>

Covering more than 20% of the earth land surface, and home to about 40% of the world's 7.4 billion inhabitants,<sup>7</sup> countries of South and Southeast Asia and LA exhibit a striking heterogeneity in terms of sociopolitical, economic and developmental indices and variations in access to products and services, including healthcare. The governmental healthcare spending in most countries of these regions is low compared to countries in North America and Western Europe.<sup>1,3</sup> Private hospitals providing sophisticated care co-exist with severe lack of public sector health infrastructure, including deficiency of primary care.<sup>8</sup>

Disease epidemiology in Asia and LA, including that of AKI, reflects these inequalities. Characteristics of patients with AKI encountered in tertiary care hospitals in large cities are similar to their counterparts in high-income countries: most are elderly, critically ill with multiorgan failure, have chronic comorbidities, and the main AKI causes are ischemia, sepsis and nephrotoxic drugs. By contrast, AKI is frequently encountered in the community in smaller urban areas distant from large cities and in rural zones secondary to diarrhea, tropical infectious diseases, animal venoms, natural medicines and poor obstetric care, amongst previously healthy young individuals.<sup>8-15</sup> Most patients present late to the healthcare system, with advanced stage disease. Access to renal replacement therapy (RRT) is variable, due either to non-availability of resources or lack of affordability, and is not rare that a RRT

cannot be delivered despite being indicated.<sup>8,14,15</sup> There is paucity of epidemiological studies on this kind of community-acquired AKI, but recent meta-analyses have shown a high frequency and high early mortality for these patients.<sup>8,15</sup> The long-term consequences of surviving a community-acquired AKI, including morbidity, mortality, effects on quality of life and economic burden, are virtually unknown.

In this review, we will focus on the epidemiology, clinical picture, prevention, risk factors and pathophysiology of AKI associated with some important tropical diseases and animal venom in tropical regions of Asia and LA, and discuss potential challenges due to emerging issues.

## **Tropical infectious diseases-associated AKI**

Despite systematic efforts at containment, infectious diseases continue to be important contributors to the burden of AKI in the tropics. Tropical AKI closely tracks the state of public health services, especially inadequate sanitary infrastructure, lack of preventive measures and poor access to health care. An estimate of the numbers affected is thwarted due to poorly organized systems of surveillance. Since patterns of organ involvement are not tracked, estimates of frequency of kidney involvement are either rough approximations or skewed towards those with advanced stage disease, since those patients are preferentially referred to tertiary care hospitals. Besides the high mortality and morbidity, these conditions impose large economic burden on countries with already limited resources. The escalating rate of tourism, work and business related travel and forced immigration due to poverty and wars makes it increasingly likely that pathogens and vectors once limited to a geographic area will spread to previously non-endemic regions. Tropical infections are often diagnosed late in non-endemic areas, with deleterious consequences.

A number of patients with tropical infections develop AKI in the setting of an undifferentiated febrile illness. In the absence of obvious evidence of common bacterial infections, such as respiratory or urinary tract infection, specific tropical infections need to be considered in the differential diagnosis. It is extremely important to recognize these syndromes not only because untreated infection may be fatal but also due the fact that the AKI incidence in population of febrile patients may be up to 41%. Patients can have additional manifestations related to other organ systems, such as liver, nervous system, or heart dysfunction, coagulopathy and thrombocytopenia. Identification of these syndromes helps in narrowing the list of diagnostic possibilities. The common differential diagnosis of tropical acute febrile illness based on organ involvement is summarized in Table 1.

We will discuss some important tropical infectious diseases associated to AKI below (summarized in table 2).

### *Malaria*

A mosquito-borne disease transmitted by the bite of infected females *Anopheles* mosquito, malaria is caused by protozoans of genus *Plasmodium*. The global effort in combating malaria has resulted in an 18% decline in the number of cases worldwide over the last 15 years. Still, there were 214 million cases and 438,000 deaths worldwide due to malaria in 2015. Children < 5 years accounted for about 70% of all deaths. Approximately 3.2 billion individuals in 95 countries, including all South and Southeast Asian countries and most LA countries were at risk of malaria in 2015.<sup>16,17</sup> South and Southeast Asia accounts for approximately 10% of the world burden of malaria, and had about 19.5 million cases and 29 thousand malaria-related deaths in 2015.<sup>18</sup> In the American continent, malaria is endemic in 21 countries and approximately 20% of the population is at risk of infection. In 2014, 389,390 confirmed cases and 87 malaria-associated fatalities were reported to the Pan American Health Organization. Brazil presently is responsible for 42 % of all cases reported in LA. Transmission remains entrenched in the Amazon Basin, which accounts for 99.5 % of the country's malaria load.<sup>19</sup> Severe malaria, associated with high level of parasitemia (>5 %) is associated with dysfunction of major organs, and is a medical emergency.<sup>11</sup>

AKI develops following infection with *P falciparum*, *P vivax* and *P knowlesi* and more rarely after *P. ovale* infection.<sup>20-24</sup> The frequency varies from 1 to 4% in all patients, and goes up to 60% among those with severe disease.<sup>20,25</sup> Recent data suggest change in disease behavior, with a rise in the frequency of malaria-associated AKI from 4.7% in 1983-95 to 17% in 1996-2008 in certain regions.<sup>26</sup> Older reports described AKI as a complication of *falciparum* infection, but recently AKI has been increasingly reported in association with *vivax* in SA and LA and *knowlesi* in Southeast Asia.<sup>27</sup> While some of these discrepancies are likely due to

different AKI definitions used, possible changes in virulence of the causative protozoa must be considered. Severe parasitemia, age < 5 years, pregnancy, HIV infection, shock, hepatic involvement, acute respiratory distress syndrome (ARDS) and disseminated intravascular coagulation (DIC) have been identified as risk factors for development of malaria-associated AKI.<sup>13,23,28,29</sup> Whereas previous literature emphasizes severe AKI, with oliguria and need for RRT, cases with earlier stages of AKI are being increasingly reported.<sup>22,24</sup>

Factors implicated in the pathogenesis of malaria-associated AKI include vascular obstruction by infected erythrocytes, generation of reactive oxygen species, immune complex deposition, hypovolemia,<sup>28</sup> and systemic inflammation<sup>30,31</sup> Renal histology usually shows acute tubular necrosis (ATN), but cortical necrosis (CN), thrombotic microangiopathy, and glomerulonephritis have also been described.<sup>22,32-36</sup> AKI increases mortality risk and length of hospitalization.<sup>23</sup> Lethality might be up to 45%<sup>28</sup>, and has been associated with disease stage, severity scores, inotropics and ventilator support.<sup>22</sup> In children, DIC, jaundice and parasitemia were associated with mortality.<sup>37</sup> Prompt diagnosis, early antimalarial treatment, timely initiation of RRT and supportive therapy were associated with improved outcome and recovery of renal function in patients with malaria-induced AKI.<sup>22,38,39</sup>

### *Leptospirosis*

The world's commonest zoonosis is caused by spirochetes of the genus *Leptospira*, and is endemic in large parts of Asia and LA. Humans are infected accidentally when the organisms shed in urine by the natural vectors (rodents, dogs, pigs, cattle, horses) enter the circulation through broken skin or mucosal membranes exposed to contaminated water or soil.

Outbreaks occur after floods, hurricanes and earthquakes. Leptospirosis is an occupational hazard for farmers, pet traders, veterinarians, rodent exterminators, slaughters, garbage

collectors and sewer workers, since they are more likely to be exposed to contaminated material.<sup>40-42</sup>

The actual burden of leptospirosis is likely underestimated, since its diagnosis is often missed or mistaken due to the nonspecific nature of clinical presentation.<sup>40,43</sup> There are an estimated 1.03 million cases worldwide with 58,900 deaths annually, about half amongst young adult males (20 to 49 years). SA and tropical LA carry the highest morbidity and mortality burden.<sup>44</sup> A recent increase in disease frequency has been described in both Asia and LA.<sup>41,45</sup>

The clinical manifestations are variable, with most patients developing minor flu-like illness, headache, myalgia, nausea, vomits, skin rash and/or conjunctivitis.<sup>40-42</sup> Electrocardiographic abnormalities were described in almost 60% of hospitalized patients.<sup>46</sup> Other findings include hepatosplenomegaly, jaundice, and hemorrhages. Weil syndrome is the name attached to its most severe multisystemic presentation that includes ARDS, diffuse alveolar bleeding, pulmonary edema, hepatic failure and AKI.<sup>40,41</sup> Kidney involvement is practically universal, since the spirochetes infiltrate renal tissue. Urinary electrolyte wasting can lead to hypokalemia and hypomagnesemia.<sup>40,41,47</sup> AKI, usually non-oliguric and frequently accompanied by hypokalemia, is seen in 40 - 87% cases. Tubular disorders precede the decline in glomerular filtration rate (GFR).<sup>48-50</sup> Data on risk factors for AKI are scarce. In an outbreak in Philippines, duration of illness before admission, elevated baseline serum creatinine and thrombocytopenia were independent risk factors for AKI.<sup>42</sup> A multicenter study showed that urinary and plasma NGAL levels were elevated in leptospirotic patients who developed AKI as compared to those with preserved renal function.<sup>51</sup> AKI etiopathogenesis is multifactorial, including direct action of the bacteria on renal tissue, hypovolemia, hypotension, rhabdomyolysis, hyperbilirubinemia, and spirochete membrane proteins-induced glycocalyx and endothelial injury<sup>40,51-53</sup> Tubular dysfunction has been related to inner medullary collecting duct vasopressin resistance, proximal tubular sodium

and water tubular transporters and  $\alpha$ -Na-K-ATPase decreased expression, and enhanced Na-K-2Cl cotransporter expression.<sup>41,54</sup> Renal histology shows interstitial nephritis, ATN and vasculitis.<sup>40,54</sup> Presence and severity of AKI and oliguria are associated with mortality<sup>43,49,55-57</sup>. Data on long-term outcome after AKI are rare. A study from Sri Lanka found that 9% of patients had developed stage 3 chronic kidney disease (CKD) one year after an episode of leptospirosis-associated AKI.<sup>58</sup> A community-based study showed that individuals with previous leptospira exposure had lower GFR and a higher frequency of CKD than those without leptospira exposure. Multivariable analyses confirmed the association between leptospiral infection and lower GFR.<sup>59</sup> A study from Brazil assessing patients with leptospirosis-associated AKI found that the creatinine clearance recovered three months after discharge, but the abnormal fractional excretion of potassium and urinary concentration capacity persisted. After six months, only urinary concentration ability remained impaired.<sup>60</sup> The mainstay of treatment is antibiotics of the penicillin group, but azithromycin, chloramphenicol, doxycycline, and cephalosporins have also been used. Patients can recover spontaneously with adequate supportive care, including fluid management and ventilatory support, and the actual benefits of antibiotics in leptospirosis remain unproved.<sup>40,41,61,62</sup> Early protective ventilatory support, pulse of corticosteroids and extra corporeal membrane oxygenation have been anecdotally used in severe leptospirosis complicated by pulmonary hemorrhage.<sup>63-65</sup> Early daily hemodialysis reduced mortality of severe leptospirosis in a series of Brazilian patients.<sup>66</sup>

### *Dengue*

A viral disease caused by RNA arbovirus of the genus *Flavivirus* (family *Flaviviridae*), dengue is transmitted by female mosquitos of the genus *Aedes*, *aegypti* and *albopictus*. Four serotypes of dengue virus cause the disease and a new serotype was recently discovered in Malaysia.<sup>67-69</sup> Dengue has emerged as a worldwide public health problem, and is an example

of global dissemination of the vectors and their ability to adapt in domestic environments secondary to increased human mobility, climate changes and the unplanned urban growth.<sup>67,68</sup> Recent studies assessed the number of people at risk at about 4 billion in 128 countries,<sup>70</sup> and put the number of annual infections at 390 million, more than three times the World Health Organization (WHO) estimate,<sup>71</sup> with an projected annual cost of US\$8.9 billion.<sup>72</sup> Southeast Asia records about 2.9 million dengue episodes and 5,906 deaths annually, with a yearly monetary burden of \$950 million.<sup>73</sup> The number of dengue cases in India doubled from 2014 to 2015, with almost 10-fold increase in some states.<sup>74</sup> Similar increases have been reported from Pakistan.<sup>75</sup> In 2015, South America had 2.3 million suspected cases, with 10,276 severe cases and 1,181 deaths.<sup>76</sup> As of April 2015, the Brazilian Ministry of Health had registered 1,254,907 notified cases of dengue, accounting for most of the disease burden in LA<sup>77</sup>. A probabilistic sensitivity analysis estimated that Mexico had about 139,000 symptomatic and 119 fatal cases annually (2010-2011), with an annual cost of \$87 million related to the illness and a yearly disease burden of 65 disability-adjusted life years per million population.<sup>78</sup>

Dengue can be asymptomatic or present as a subclinical infection. The clinical forms of the disease are classified as dengue fever, dengue hemorrhagic fever without shock or with shock and expanded dengue syndrome.<sup>79,80</sup> Dengue fever (DF) is an acute illness, characterized by fever, headache, retro-orbital pain, myalgia, joint pain, nausea and vomiting, itchy cutaneous rash and, occasionally hemorrhagic manifestations. Secondary infection, i.e. a re-infection with a different virus serotype from the primary infection, can result in severe forms of the disease. These forms are characterized by thrombocytopenia, evidence of capillary leak, such as elevated hematocrit, hypoalbuminemia, serous effusions and by hemodynamic instability.

Renal involvement can manifest as AKI, dyselectrolytemias, proteinuria, hematuria, and hemolytic uremic syndrome.<sup>68,81-88</sup> AKI frequency varies from 0.9% in Thai children<sup>89</sup> to

30.7 % in Southern Indian adults.<sup>90</sup> Studies using new AKI criteria reported frequencies up to 14.2% by AKIN and 12.6% by RIFLE.<sup>91,92</sup> AKI has been described in all forms of dengue<sup>93</sup> and the risk factors include elevated liver enzymes, low serum albumin, decreased serum bicarbonate, coexisting bacterial or viral infection, sepsis, multiple organs dysfunction, inotropic drugs, older age, obesity, severity of dengue infection, rhabdomyolysis, diabetes mellitus, late hospitalization and nephrotoxic drugs.<sup>92,94</sup> Postulated pathogenetic factors include direct viral effects on renal tissue, hemodynamic instability, rhabdomyolysis, hemolysis, acute glomerular injury, and systemic inflammatory response syndrome.<sup>68,94-97</sup> Histology showed ATN, thrombotic microangiopathy and glomerulopathy.<sup>98-101</sup> AKI is consistently associated with longer hospital stay and increased mortality.<sup>94,102-104</sup> Management entails careful volume replacement and supportive care depending upon manifestations.<sup>105</sup> The use of parenteral corticosteroids for severe dengue is debatable, and recommendations for its use in AKI patients does not exist.<sup>106</sup> It is advisable to monitor serum creatinine-kinase phosphate levels for early diagnosis of rhabdomyolysis and institution of appropriate preventive measures.

### *Scrub typhus*

Scrub typhus is caused by the bacterium *Orientia tsutsugamushi* (formerly *Rickettsia tsutsugamushi*), and is transmitted to humans by infected *Leptotrombidium* mites larva (chiggers). An estimated 1 billion people worldwide are at risk, and an estimated 1 million cases occur each year.<sup>107</sup> The disease is endemic in South and Southeast Asia.<sup>108</sup> There is only one documented case reported in South America.<sup>109</sup> Manifestations range from mild, asymptomatic disease to a severe, fatal illness. Findings include lymphadenopathy, skin rash, fever, myalgia, headache, gastrointestinal symptoms and cough. A typical eschar occurs in the site of the bite, but is not always present. Severe disease is characterized by ARDS, AKI, bleeding, coagulation disorders, meningoencephalitis and shock.<sup>110-115</sup>

Renal involvement is common. Urinary abnormalities such as hematuria, proteinuria, pyuria and presence of granular casts occur in 50-80% patients.<sup>116,117</sup> AKI is frequent, ranging from 20% to up to 60%.<sup>90,100,112-119</sup> The possible mechanisms of AKI are infiltration and direct effects of the bacteria on kidney tissue<sup>120,121</sup>, intravascular hemolysis associated to glucose-6-phosphate dehydrogenase-deficiency<sup>122</sup>, rhabdomyolysis<sup>116,123,124</sup> renal ischemia due to hemodynamic instability and vasculitis.<sup>116</sup> Histopathology shows ATN, interstitial nephritis and mild mesangial glomerulonephritis.<sup>120,125-127</sup> AKI has been consistently associated to higher short-term mortality.<sup>115-117</sup> Early and adequate administration of doxycycline promotes remarkable improvement in the majority of patients, including renal function recovery.<sup>113,128</sup>

### *Yellow fever*

Yellow fever (YF) is caused by the YF virus (YFV), an arbovirus considered the prototype for the *Flavivirus* genus and *Flaviviridae* family, which is endemic in areas of Africa and South America. The vectors are blood-feeding mosquitos of the genera *Haemagogus* in South America and *Aedes* in Africa. Massive vaccinations practically eliminated the disease in North America and Europe, and reduced considerably its incidence in Central and South America and Africa. However, due the existence of an YFV cycling between forest mosquitos and wild non-human primate hosts in Africa and Americas, YF has not been totally eradicated. Thirty-four countries in Africa and 13 countries in Central and South America have areas endemic for YF. The burden of YF during 2013 was estimated as 84,000 to 170,000 severe cases and 29,000 to 60,000 deaths. Practically 90% of YF cases and deaths occur in Africa. The dissemination of the urban mosquito vector, *Aedes aegypti*, and the escalation in air travel brings the fear of the introduction of the YFV outside endemic regions, particularly in the unimmunized densely populated areas of Southeast Asia, as recently seen with the worldwide dissemination of Chikungunya and Zika virus.<sup>129-133</sup> YF might range from an asymptomatic or mild illness to a fulminating, life threatening, multiorgan disease. A large

number of cases experience a self-limited disease, characterized by high fever, bradycardia, chills, anorexia, headache, muscle and back pain and prostration. Fifteen to 25% of the patients develop the most severe form of the disease, with hepatic failure, jaundice, hemorrhagic diathesis, epigastric pain, vomits, shock and AKI. Death occurs in up to 50% of these cases. Survivors experience a long convalescent phase, with weakness and fatigue.<sup>129-131, 134,135</sup>

Although AKI is always mentioned as part of severe YF<sup>129-131,135,136</sup> information about the frequency, risk factors, and etiopathogenesis of YF-associated AKI are very scarce. Oliguria ensued after five to seven days in severe YF, and even earlier in African patients without jaundice or hepatic abnormalities.<sup>137,138</sup> Renal histology disclosed ATN.<sup>135,139</sup> The mechanisms causing AKI are obscure. Renal ischemia, intravascular coagulation, shock, bilirubin-induced tubular toxicity, YF virus direct effect on the kidneys and an enormous inflammatory cytokines release are possible mechanisms of AKI.<sup>140-145</sup>

### **Animal venom-induced AKI**

Accidents caused by venomous animals, such as snakes and arthropods (bees, wasps, spiders and caterpillars) are a significant public health issue in several areas of tropical countries. AKI is one of the main effects of these venoms, and associated with high morbidity and lethality. We will review below the most relevant aspects of AKI associated to snakebite (Russell's viper, Bothrops and Crotalus) and venomous arthropods (bees, Lonomia caterpillars, Loxosceles spiders and scorpions) in SA and LA (summarized in table 3).

#### *Snake Bite*

Like several tropical infections, venomous snake bites have been categorized as a neglected public health issue, particularly in LA, Asia and Africa by the WHO.<sup>146</sup> Data on the global burden of human envenomation due to snakebite estimates 421,000 to 1,841,000 cases with

20,000 to 94,000 deaths per year. SA and LA contribute the largest number of cases and SA had the higher mortality.<sup>147</sup> Vulnerable populations, such as the economically disadvantaged and those living in rural communities are affected disproportionately.<sup>148</sup> Because the available epidemiological information is almost completely based on hospital data and likely to have missed victims treated in community healthcare systems, the actual numbers are likely much higher.<sup>149</sup>

Snake venoms have a complex chemical composition, consisting mostly of potentially toxic proteins and enzymes, carbohydrates, lipids, metals, biogenic amines, free amino acids and nucleotides. They have evolved evolutionarily as hunting weapons, and human bites are accidental.<sup>150</sup> Clinical manifestations depend on the type and inoculated venom dose, bite site, victim body size and medical intervention timeliness. They range from mild local symptoms to severe systemic envenoming, which may become apparent in a few hours.

AKI is one of the most important causes of death in patients bitten by snakes of the class Viperidae and Hydrophidae.<sup>151,152</sup> The majority of AKI cases is caused by Russell's viper accidents in Asia and by snakes of the genus Bothrops and Crotalus in LA.<sup>153-158</sup> Snake venoms are concentrated in renal tissue within minutes and appear in urine within a few hours after inoculation.<sup>159-161</sup> Unless preventive measures are implemented promptly, early renal dysfunction occurs frequently (see table 2).<sup>157,162-168</sup> AKI can be severe, oliguric and dialysis dependent.<sup>10,162,169,170</sup> Associated clinical findings might include, depending on the offending snake, those related to local injury (figure 1), rhabdomyolysis, intravascular hemolysis, consumptive coagulopathy, myocardial injury, neurotoxicity, loin pain, hypopituitarism and systemic inflammatory response syndrome.<sup>154,157,169-177</sup> Although most patients recover renal function, AKI after snakebite has been associated with development of CKD in both adults and children.<sup>178-180</sup> and may aggravate renal dysfunction in agrochemical nephropathy.<sup>181</sup>

The most frequent renal histological abnormality found in snake venom-associated AKI is ATN.<sup>169,182-185</sup> A small but significant minority of cases develop acute CN or acute interstitial nephritis (AIN). CN has been described mainly following Russell's viper and Bothrops bites<sup>182,184,186-188</sup>, and foresees a bleak renal prognosis. Bilateral and diffuse involvement is associated with irreversible renal failure. CN should be suspected when oliguria persists beyond two weeks and can be confirmed by renal biopsy or a contrast-enhanced CT scan (figure 2). Glomerular injury has also been described, and is manifested histologically as ballooning of glomerular capillaries, splitting of glomerular basement membrane, swelling of endothelial cells, and focal mesangial cell proliferation.<sup>189</sup>

Several pathogenetic factors contribute to AKI, including hemodynamics abnormalities, hemolysis, rhabdomyolysis, direct tubular toxicity, systemic inflammation, activation of coagulation pathway leading to glomerular deposition of microthrombi, oxidative stress, hyperuricemia, release of cytochrome C and induction of apoptosis in tubular epithelium.<sup>170,183,185,190-203</sup>

The most effective way to prevent AKI is early and adequate administration of antivenom<sup>204</sup>. A prospective study on AKI after *Crotalus* snakebite in Brazil showed that an interval of over two hours between the bite and antivenom administration raised the AKI risk 10-fold after correction for other factors.<sup>162</sup> Similar association between delay in the timing for receiving antivenom and AKI have been reported in retrospective studies after Bothrops and Russell's viper bites.<sup>205-207</sup> In experiments with fresh isolated proximal tubules antivenom only prevent tubular injury when added to the median simultaneously with the venom.<sup>190</sup> Limited availability and perceived risk of side effects have prompted to research in alternative antivenom therapies, none of whom have shown any promise.<sup>208,209</sup> The available antivenom in Asia is usually a polyvalent form raised against the "Big Four" (*D. russelii*, *Naja naja*,

*Echis carinatus* and *Bungarus caeruleus*), which may lead to incomplete neutralization of toxins of Russell's viper venom.<sup>210-212</sup>

### *Bees*

Honeybees, hornets and yellow jackets use venom as a defensive weapon aimed to protect the hive against intruders. Venom components are melittin (50%), histamine, hyaluronidase, apamin, acid phosphatase and phospholipase A<sub>2</sub> A<sub>2</sub>.<sup>213,214</sup> Few stings usually cause only pain and allergic reactions. However, multiple stings following attack by a swarm introduces large doses of venom into circulation, causing severe multisystemic effects, including hemolysis, rhabdomyolysis, coagulation, cardiac, pulmonary, hepatic and nervous system disorders, hypertension and AKI.<sup>214- 218</sup> Of particular note are the Africanized honeybees found in LA, generated because of interbreeding between African honeybees that escaped from a Brazilian facility and European honeybees. They are much more aggressive than their predecessors and caused a spike in the number of cases in the region.<sup>214,219</sup>

Cases of AKI associated with honeybee attacks have been reported in India<sup>220-222</sup> and LA.<sup>219,223-231</sup> The largest series describes 43 patients with Africanized honeybee venom-associated AKI in Colombia. The victims were typically men from rural areas, with approximately 900 stings per individual. Almost all presented with rhabdomyolysis and oliguric dialysis-dependent AKI, which developed within 48 hr. Mortality was 16.3%.<sup>229</sup> The probable mechanisms of AKI are renal vasoconstriction, direct proximal tubular cells toxicity, hemolysis, oxidative stress, systemic release of inflammatory cytokines and pigmenturia.<sup>214,232,233</sup> Experimental studies showed that the injury can be mitigated by the use of human antibody fragments binding melittin and phospholipase A<sub>2</sub>.<sup>234</sup> The most frequent renal histology finding is ATN.<sup>213,219,222,232,235</sup> The morbidity and lethality of massive attacks has prompted the development of a hyperimmune equine serum antivenom against

Africanized honeybee venom, which was effective in protecting against rhabdomyolysis and hemolysis in mice; it is yet unknown if this antivenom will prevent AKI.<sup>236</sup> Plasmapheresis, aiming to remove venom and inflammatory mediators, was used with success in a patient with more than 2,000 Africanized honeybees stings.<sup>231</sup> CVVH has been used in patients with honeybee venom-associated rhabdomyolysis and AKI, and led to decrease in serum myoglobin levels in 24 hours.<sup>237</sup>

### *Lonomia caterpillars*

Caterpillars of the genus *Lonomia* have as natural habitat the trees in the deep forests of LA. They have a body covered by bristles, which inject venom when the victim touches the caterpillar (figure 3).<sup>213,238</sup> Deforestation have caused migration of these animals close to human habitations, increasing the number of accidents.<sup>239,240</sup> The clinical picture of these accidents is characterized by burning at the contact site, headache, fever, vomiting and a bleeding diathesis, which might be life threatening.<sup>213,241</sup>

*Lonomia* venom-associated AKI has been reported mainly in Brazil.<sup>242-245</sup> Early antivenom administration in rats prevented *Lonomia* venom-induced hemorrhagic manifestations and increase in serum creatinine.<sup>246</sup> The probably mechanisms of AKI are direct nephrotoxicity, intravascular hemolysis, systemic and endothelial inflammatory pathway activation, hypotension, increased renal tissue expression of proteins involved in cell stress, inflammation, heme-induced oxidative stress, coagulation and complement system activation.<sup>247-249</sup> Renal histology have shown ATN, glomerular fibrin deposition and ischemia and tubular atrophy.<sup>243,244,249</sup> Development of CKD following *Lonomia* venom-associated AKI was described in five patients.<sup>245,250</sup>

## *Loxosceles*

Spiders of the genus *Loxosceles* are distributed worldwide, including SA and LA. Five species (*Loxosceles rufescens*, *L. laeta*, *L. intermedia*, *L. gaucho* and *L. reclusa*) are mostly responsible for human envenomation, known as loxoscelism, a major public health problem in South America (especially in Brazil, Chile and Peru), and also described in India.<sup>251,252</sup>

*Loxosceles* are well adapted to domestic environment, where they hide inside clothes, towels and bed sheets and bite the victim upon contact.<sup>251,252</sup> The bite may go unnoticed, and the first presentation (12-24 h after the bite) takes the form of painful indurated edema, erythema, ecchymosis and hemorrhagic blister, which frequently progresses slowly (72 hours) to local necrosis and ulceration (figure 4). In approximately half of the cases, the cutaneous injury progresses to a dry necrotic round scar with defined margins. About half the patients exhibit systemic features, such as rash, fever, pruritus, malaise, headache, nausea and vomit.<sup>251,252</sup> Up to 27% of the patients develop viscerocutaneous or systemic loxoscelism, which is more frequent in Peru, Chile and some areas of South Brazil, characterized by intravascular hemolysis, jaundice, coagulation disorders, thrombocytopenia, rhabdomyolysis, hypotension/shock and AKI.<sup>213,251-254</sup>

*Loxosceles* venom-associated AKI was described rarely in India.<sup>255,256</sup> In Brazil AKI was reported ranging from 3.3 to 6.4%.<sup>254,257</sup> Cases were also described in Chile<sup>258,259</sup> and Peru.<sup>260</sup> The risk factors associated with AKI are severity of hemolysis, and accidents caused by *L. laeta*. The mechanism of AKI might be linked to direct venom nephrotoxicity, hemoglobinuria, myoglobinuria, renal vasoconstriction, and systemic inflammation.<sup>213,253,257,261-263</sup> Renal histology have shown pigment-induced ATN.<sup>258,263,264</sup> Antivenom is probably the most effective therapy against AKI, but its application is often delayed as the bite goes unnoticed.<sup>265</sup> There is one case report relating a successful treatment with plasma exchange in a child with severe systemic loxoscelism, including AKI.<sup>266</sup>

## Scorpions

Scorpions are venomous arthropods, responsible for over one million of accidents worldwide.<sup>267</sup> Severe envenomation is a public health issue in parts of LA, South Asia, North Africa, and Middle East,<sup>268,269</sup> and in some countries, such Brazil, scorpion stings exceed the number of cases of snakebite.<sup>270</sup> The genera *tityus* (South America) and *centruroides* (Mexico and Central America) in LA and the *mesobuthus* in Asia (particularly India) are medically relevant.<sup>268,270,271</sup> Scorpion venom is a hunting and defensive weapon, composed by several toxins. The toxins responsible for most of the medically relevant effects of the venom are small polypeptides ( $\alpha$ -toxins), which inhibit mammalian voltage-gated sodium channels causing prolonged depolarization.<sup>268,272</sup> Scorpion venom causes autonomic overactivity, and a massive release of vasoactive substances and inflammatory cytokines.<sup>268,272</sup> The clinical picture is dominated by signs and symptoms of excessive cholinergic and adrenergic activity, cardiac dysfunction, and neuromuscular excitation. Severe cases, more common in children, may evolve to pulmonary edema, multiorgan failure and death.<sup>268-270,272,273</sup> Stings by the *H. lepturus* in Iran cause cytotoxic features, including hemolysis.<sup>268,</sup>

The venom is cleared mainly by the kidneys and liver and concentrates rapidly and intensively in renal tissue.<sup>274</sup> However, scorpion venom-associated AKI is rarely reported, with the majority of cases described in Middle East, and few cases in LA and SA.<sup>269,271,275,276</sup> The mechanisms of renal injury are likely renal ischemia due to vasoconstriction, hemodynamic instability, rhabdomyolysis, excessive systemic inflammation, and direct venom nephrotoxicity.<sup>269,271,272,277</sup> Renal histology shows ATN, glomerular changes, interstitial infiltrate, TMA and CN.<sup>269,272,276-279</sup> A case-control study in Mexico suggested that scorpion accidents were associated with development of CKD in children.<sup>280</sup> Use of

antivenom partially decreased kidney venom concentration in a mice model, and has been associated with more favorable clinical outcome.<sup>268,274</sup>

### **Future challenges**

Tropical regions of Asia and LA are likely to face major challenges to kidney health because of climate change and water scarcity. According to the U.K.-based risk analysis firm Maplecroft, the top 10 countries at “extreme risk” from climate change are all tropical countries. Kidneys are likely to be particularly vulnerable to heat stress, and the predicted re-emergence of water- and vector-borne infectious diseases. There is evidence that virulence of disease-causing organisms is changing, as noted by emergence of kidney injury in *vivax* malaria and scrub typhus and control getting harder due to emergence of antimicrobial resistance. Degradation of ecosystem, air and water pollution will increase the risk of exposure to venomous snakes and insects that were hitherto limited to areas away from human inhabitation. This unexpected burden of tropical diseases will likely further stress an overloaded healthcare system leading to deterioration in outcomes. We need more research in order to better understand the impact of infection and venom associated AKI on the long-term risk of CKD. This is of particular relevance as an increasing number of CKD cases are described in relatively young individuals where the etiology cannot be determined.

## References

1. Jha V. ESRD burden in South Asia: the challenges we are facing. *Clin Nephrol.* 2015;83(7 Suppl 1):7-10.
2. Worldbank data. <http://data.worldbank.org/region/SAS> accessed on June 2, 2016.
3. Cusumano AM, Garcia-Garcia G, Gonzalez-Bedat MC, et al. Latin American Dialysis and Transplant Registry: 2008 prevalence and incidence of end-stage renal disease and correlation with socioeconomic indexes. *Kidney Int Suppl.* (2011); 2013;3:153-156.
4. Rosa-Diez G, Gonzalez-Bedat M, Pecoits-Filho R, et al. Renal replacement therapy in Latin American end-stage renal disease. *Clin Kidney J.* 2014;7:431-436.
5. "World Population Prospects, The 2015 Revision: Key Findings and Advance Tables". United Nations Department of Economic and Social Affairs, Population Division. July 29, 2015. pp. 13–17.
6. Worldbank data. <http://data.worldbank.org/region/LAC> accessed on June 2, 2016.
7. Worldmeters data. <http://www.worldometers.info/world-population> accessed June 2, 2016.
8. Mehta RL, Cerdá J, Burdmann EA, et al. International Society of Nephrology's 0by25 initiative for acute kidney injury (zero preventable deaths by 2025): a human rights case for nephrology. *Lancet.* 2015;385:2616-43.
9. Santos WJ, Zanetta DM, Pires AC, et al. Patients with ischaemic, mixed and nephrotoxic acute tubular necrosis in the intensive care unit--a homogeneous population? *Crit Care.* 2006;10:R68.

10. Lombardi R, Yu L, Younes-Ibrahim M, et al. Epidemiology of acute kidney injury in Latin America. *Semin Nephrol.* 2008;28:320-9.
11. Jha V, Chugh KS. Community-acquired acute kidney injury in Asia. *Semin Nephrol.* 2008;28:330-47.
12. Cerdá J, Bagga A, Kher V, et al. The contrasting characteristics of acute kidney injury in developed and developing countries. *Nat Clin Pract Nephrol.* 2008;4:138-53.
13. Jha V, Parameswaran S. Community-acquired acute kidney injury in tropical countries. *Nat Rev Nephrol.* 2013;9:278-90.
14. Lombardi R, Rosa-Diez G, Ferreiro A, et al. Acute kidney injury in Latin America: a view on renal replacement therapy resources. *Nephrol Dial Transplant.* 2014;29:1369-76.
15. Mehta RL, Burdmann EA, Cerdá J, et al. Recognition and management of acute kidney injury in the International Society of Nephrology 0by25 Global Snapshot: a multinational cross-sectional study. *Lancet.* 2016;387:2017-25.
16. World Malaria Report 2015 WHO.
17. WHO page <http://www.who.int/mediacentre/factsheets/fs094/en/> accessed on June 16 2016
18. Regmi K, Kunwar A, Ortega L. A systematic review of knowledge, attitudes and beliefs about malaria among the South Asian population. *Infect Ecol Epidemiol.* 2016;6:30822.
19. Ferreira MU, Castro MC. Challenges for malaria elimination in Brazil. *Malar J.* 2016;15:284.
20. Naqvi R, Ahmad E, Akhtar F, et al. Outcome in severe acute renal failure associated with malaria. *Nephrol Dial Transplant.* 2003;18:1820–23.

21. Prakash J, Singh AK, Kumar NS et al. Acute renal failure in Plasmodium vivax malaria. J. Assoc. Physicians India. 2003;51:265–7.
22. Kute VB, Shah PR, Munjappa BC, et al. Outcome and prognostic factors of malaria-associated acute kidney injury requiring hemodialysis: A single center experience. Indian J Nephrol. 2012;22:33-8.
23. Saravu K, Rishikesh K, Parikh CR. Risk factors and outcomes stratified by severity of acute kidney injury in malaria. PLoS One. 2014;9:e90419.
24. Naqvi R. Plasmodium Vivax causing acute kidney injury: A foe less addressed. Pak J Med Sci. 2015;311:472-5
25. Mehta KS, Halankar AR, Makwana PD, et al. Severe acute renal failure in malaria. J Postgrad Med. 2001;47:24-6.
26. Prakash J, Singh TB, Ghosh B, et al. Changing epidemiology of community-acquired acute kidney injury in developing countries: analysis of 2405 cases in 26 years from eastern India. Clin Kidney J. 2013;6:150-5.
27. Kaushik R, Kaushik RM, Kakkar R, et al. Plasmodium vivax malaria complicated by acute kidney injury: experience at a referral hospital in Uttarakhand, India. Trans R Soc Trop Med Hyg. 2013;107:188-94.
28. Mishra SK, Das BS. Malaria and acute kidney injury. Semin Nephrol. 2008;28:395-408.
29. Conroy AL, Hawkes M, Elphinstone RE, et al. Acute Kidney Injury Is Common in Pediatric Severe Malaria and Is Associated With Increased Mortality. Open Forum Infect Dis. 2016;3:ofw046.

30. Herbert F, Tchitchek N, Bansal D, et al. Evidence of IL-17, IP-10, and IL-10 involvement in multiple-organ dysfunction and IL-17 pathway in acute renal failure associated to Plasmodium falciparum malaria. *J Transl Med.* 2015;13:369.
31. Plewes K, Royakkers AA, Hanson J, et al. Correlation of biomarkers for parasite burden and immune activation with acute kidney injury in severe falciparum malaria. *Malar J.* 2014;13:91.
32. Sinha A, Singh G, Bhat AS, et al. Thrombotic microangiopathy and acute kidney injury following vivax malaria. *Clin Exp Nephrol.* 2013;17:66-72.
33. Kanodia KV, Vanikar AV, Kute VB, et al. Plasmodium vivax malaria associated with acute post infectious glomerulonephritis. *Ren Fail.* 2013;35:1024-6.
34. Patel MP, Kute VB, Gumber MR, et al. An unusual case of Plasmodium vivax malaria monoinfection associated with crescentic glomerulonephritis: a need for vigilance. *Parasitol Res.* 2013;112:427-30.
35. Kumar R, Bansal N, Jhorawat R, et al. Renal cortical necrosis: A rare complication of Plasmodium vivax malaria. *Indian J Nephrol.* 2014;24:390-3.
36. Nayak KC, Kumar S, Gupta BK, et al. Clinical and histopathological profile of acute renal failure caused by falciparum and vivax monoinfection: an observational study from Bikaner, northwest zone of Rajasthan, India. *J Vector Borne Dis.* 2014;51:40-6.
37. Prasad R, Mishra OP. Acute Kidney Injury in Children with Plasmodium falciparum Malaria: Determinants for Mortality. *Perit Dial Int.* 2016;36:213-7.
38. Koh KH, Chew PH, Kiyu A. A retrospective study of malaria infections in an intensive care unit of a general hospital in Malaysia. *Singapore Med J.* 2004;45:28-36.

39. Zinna S, Vathsala A, Woo KT. A case series of falciparum malaria-induced acute renal failure. *Ann Acad Med Singapore*. 1999;28:578-82.
40. Abdulkader RC, Silva MV. The kidney in leptospirosis. *Pediatr Nephrol*. 2008;23:2111-20.
41. Andrade L, de Francesco Daher E, Seguro AC. Leptospiral nephropathy. *Semin Nephrol*. 2008;28:383-94.
42. Mendoza MT, Roxas EA, Ginete JK, et al. Clinical profile of patients diagnosed with leptospirosis after a typhoon: a multicenter study. *Southeast Asian J Trop Med Public Health*. 2013;44:1021-35.
43. Varma MD, Vengalil S, Vallabhajosyula S, et al. Leptospirosis and dengue fever: a predictive model for early differentiation based on clinical and biochemical parameters. *Trop Doct*. 2014;44:100-2.
44. Costa F, Hagan JE, Calcagno J, et al. Global Morbidity and Mortality of Leptospirosis: A Systematic Review. *PLoS Negl Trop Dis*. 2015;9:e0003898.
45. Sethi S, Sharma N, Kakkar N, et al. Increasing trends of leptospirosis in northern India: a clinico-epidemiological study. *PLoS Negl Trop Dis*. 2010;4:e579.
46. Škerk V, Markotić A, Puljiz I, et al. Electrocardiographic changes in hospitalized patients with leptospirosis over a 10-year period. *Med Sci Monit*. 2011;17:CR369-75.
47. Khositseth S, Sudjaritjan N, Tananchai P, et al. Renal magnesium wasting and tubular dysfunction in leptospirosis. *Nephrol Dial Transplant*. 2008;23:952-8.
48. Abdulkader RC, Seguro AC, Malheiro PS, et al. Peculiar electrolytic and hormonal abnormalities in acute renal failure due to leptospirosis. *Am J Trop Med Hyg*. 1996;54:1-6.

49. Daher EF, Lima RS, Silva Júnior GB, et al. Clinical presentation of leptospirosis: a retrospective study of 201 patients in a metropolitan city of Brazil. *Braz J Infect Dis*. 2010;14:3-10.
50. Seguro AC, Andrade L. Pathophysiology of leptospirosis. *Shock*. 2013;39 Suppl 1:17-23.
51. Srisawat N, Praditpornsilpa K, Patarakul K, et al. Neutrophil Gelatinase Associated Lipocalin (NGAL) in Leptospirosis Acute Kidney Injury: A Multicenter Study in Thailand. *PLoS One*. 2015;10:e0143367.
52. Coursin DB, Updike SJ, Maki DG. Massive rhabdomyolysis and multiple organ dysfunction syndrome caused by leptospirosis. *Intensive Care Med*. 2000;26:808-12;
53. Libório AB, Braz MB, Seguro AC, et al. Endothelial glycocalyx damage is associated with leptospirosis acute kidney injury. *Am J Trop Med Hyg*. 2015;92:611-6.
54. Araujo ER, Seguro AC, Spichler A, et al. Acute kidney injury in human leptospirosis: an immunohistochemical study with pathophysiological correlation. *Virchows Arch*. 2010;456:367-75.
55. Taylor AJ, Paris DH, Newton PN. A Systematic Review of the Mortality from Untreated Leptospirosis. *PLoS Negl Trop Dis*. 2015;9:e0003866.
56. Daher EF, Silva GB Jr, Karbage NN, et al. Predictors of oliguric acute kidney injury in leptospirosis. A retrospective study on 196 consecutive patients. *Nephron Clin Pract*. 2009;112:c25-30.
57. Silva Júnior GB, Abreu KL, Mota RM, et al. RIFLE and Acute Kidney Injury Network classifications predict mortality in leptospirosis-associated acute kidney injury. *Nephrology (Carlton)*. 2011;16:269-76.

58. Herath NJ, Kularatne SA, Weerakoon KG, et al. Long term outcome of acute kidney injury due to leptospirosis? A longitudinal study in Sri Lanka. *BMC Res Notes*. 2014;7:398.
59. Yang HY, Hung CC, Liu SH, et al. Overlooked Risk for Chronic Kidney Disease after Leptospiral Infection: A Population-Based Survey and Epidemiological Cohort Evidence. *PLoS Negl Trop Dis*. 2015;9:e0004105.
60. Daher Ede F, Zanetta DM, Abdulkader RC. Pattern of renal function recovery after leptospirosis acute renal failure. *Nephron Clin Pract*. 2004;98:c8-14.
61. Charan J, Saxena D, Mulla S, et al. Antibiotics for the treatment of leptospirosis: systematic review and meta-analysis of controlled trials. *Int J Prev Med*. 2013;4:501-10.
62. Brett-Major DM, Coldren R. Antibiotics for leptospirosis. *Cochrane Database Syst Rev*. 2012;(2):CD008264.
63. Gomes NP, Menescal ZL, Holanda MA. Protective ventilation and alveolar recruitment maneuver in a patient with leptospirosis-induced acute respiratory distress syndrome. *J Bras Pneumol*. 2012;38:140-2.
64. Thunga G, John J, Sam KG, et al. Role of high-dose corticosteroid for the treatment of leptospirosis-induced pulmonary hemorrhage. *J Clin Pharmacol*. 2012;52:114-6.
65. Liao CY, Ben RJ, Wu HM, et al. Acute Respiratory Distress Syndrome Manifested by Leptospirosis Successfully Treated by Extracorporeal Membrane Oxygenation (ECMO). *Intern Med*. 2015;54:2943-6.
66. Andrade L, Cleto S, Seguro AC. Door-to-dialysis time and daily hemodialysis in patients with leptospirosis: impact on mortality. *Clin J Am Soc Nephrol*. 2007;2:739-44.
67. Guzman MG, Harris E. Dengue. *Lancet*. 2015;385(9966):453-65.

68. Oliveira JF, Burdmann EA. Dengue-associated acute kidney injury. *Clin Kidney J.* 2015;8:681-5.
69. Normile D. Tropical medicine. Surprising new dengue virus throws a spanner in disease control efforts. *Science* 2013;342:415.
70. Brady OJ, Gething PW, Bhatt S, et al. Refining the global spatial limits of dengue virus transmission by evidence-based consensus. *PLoS Negl Trop Dis.* 2012;6:e1760.
71. Bhatt S, Gething PW, Brady OJ, et al. The global distribution and burden of dengue. *Nature.* 2013;496:504-7.
72. Shepard DS, Undurraga EA, Halasa YA, et al. The global economic burden of dengue: a systematic analysis. *Lancet Infect Dis.* 2016 Apr 15. pii:S1473-3099(16)00146-8. doi:10.1016/S1473-3099(16)00146-8. [Epub ahead of print]
73. Shepard DS, Undurraga EA, Halasa YA. Economic and disease burden of dengue in southeast Asia. *PLoS Negl Trop Dis* 2013;7:e2055.
74. Bagcchi S. Dengue surveillance poor in India. *Lancet.* 2015;386:1228.
75. Haider Z, Ahmad FZ, Mahmood A, et al. Dengue fever in Pakistan: a paradigm shift; changing epidemiology and clinical patterns. *Perspect Public Health.* 2015;135:294-8.
76. Pan American Health Organization/WHO, Dengue Cases, Americas, 2015; <http://www.paho.org/>, accessed in June 22, 2016.
77. Fares RC, Souza KP, Añez G, Rios M. Epidemiological Scenario of Dengue in Brazil. *Biomed Res Int.* 2015;2015:321873.
78. Undurraga EA, Betancourt-Cravioto M, Ramos-Castañeda J, et al. Economic and disease burden of dengue in Mexico. *PLoS Negl Trop Dis.* 2015;9:e0003547.

79. World Health Organization. Dengue: Guidelines for Diagnosis, Treatment, Prevention and Control, New edition. Geneva: World Health Organization, 2009.
80. World Health Organization (2011). Comprehensive guidelines for prevention and control of dengue and dengue haemorrhagic fever.
81. Lima EQ, Gorayeb FS, Zanon JR, et al. Dengue haemorrhagic fever-induced acute kidney injury without hypotension, haemolysis or rhabdomyolysis. *Nephrol Dial Transplant*. 2007;22:3322-6.
82. Bhagat M, Zaki SA, Sharma S, et al. Acute glomerulonephritis in dengue haemorrhagic fever in the absence of shock, sepsis, haemolysis or rhabdomyolysis. *Paediatr Int Child Health*. 2012;32:161-3.
83. Lizarraga KJ, Nayer A. Dengue-associated kidney disease. *J Nephropathol*. 2014;3:57-62.
84. Aroor S, Kumar S, Mundkur S, et al. Hemolytic uremic syndrome associated with Dengue fever in an adolescent girl. *Indian J Pediatr*. 2014;81:1397-8.
85. Vachvanichsanong P, McNeil E. Electrolyte disturbance and kidney dysfunction in dengue viral infection. *Southeast Asian J Trop Med Public Health*. 2015;46 Suppl 1:108-17.
86. Mallhi TH, Sarriff A, Adnan AS, et al. Dengue-induced Acute Kidney Injury (DAKI): A Neglected and Fatal Complication of Dengue Viral Infection-A Systematic Review. *J Coll Physicians Surg Pak*. 2015;25:828-34.
87. Vachvanichsanong P, Thisyakorn U, Thisyakorn C. Dengue hemorrhagic fever and the kidney. *Arch Virol*. 2016;161:771-8.
88. Hebbal P, Darwich Y, Fong J, et al. Nephrotic-range proteinuria in an eight-year-old traveler with severe dengue: Case report and review of the literature. *Travel Med Infect Dis*. 2016;14:45-8.

89. Laoprasopwattana K, Pruekprasert P, Dissaneewate P, et al. Outcome of dengue hemorrhagic fever-caused acute kidney injury in Thai children. *J Pediatr* 2010;157:303-9.
90. Basu G, Chrispal A, Boorugu H, et al. Acute kidney injury in tropical acute febrile illness in a tertiary care centre-RIFLE criteria validation. *Nephrol Dial Transplant*. 2011;26:524-31.
91. Mallhi TH, Khan AH, Sarriff A, et al. Defining acute kidney injury in dengue viral infection by conventional and novel classification systems (AKIN and RIFLE): a comparative analysis. *Postgrad Med J*. 2016;92(1084):78-86.
92. Mehra N, Patel A, Abraham G, et al. Acute kidney injury in dengue fever using Acute Kidney Injury Network criteria: incidence and risk factors. *Trop Doct*. 2012;42:160-2.
93. Neeraja M, Iakshmi V, Teja VD, et al. Unusual and rare manifestations of dengue during a dengue outbreak in a tertiary care hospital in South India. *Arch Virol*. 2014;159:1567-73.
94. Mallhi TH, Khan AH, Adnan AS, et al. Incidence, Characteristics and Risk Factors of Acute Kidney Injury among Dengue Patients: A Retrospective Analysis. *PLoS One*. 2015;10:e0138465.
95. Khunchai S, Junking M, Suttitheptumrong A, et al. NF- $\kappa$ B is required for dengue virus NS5-induced RANTES expression. *Virus Res*. 2015;197:92-100.
96. Mishra A, Singh VK, Nanda S. Rhabdomyolysis and acute kidney injury in dengue fever. *BMJ Case Rep*. 2015;2015.pii:bcr2014209074.
97. Pagliari C, Simões Quaresma JA, Kanashiro-Galo L, et al. Human kidney damage in fatal dengue hemorrhagic fever results of glomeruli injury mainly induced by IL17. *J Clin Virol*. 2016;75:16-20.
98. Mohsin N, Mohamed E, Gaber M, et al. Acute tubular necrosis associated with non-hemorrhagic dengue fever: a case report. *Renal Fail* 2009;31:736-9.

99. Repizo LP, Malheiros DM, Yu L, et al. Biopsy proven acute tubular necrosis due to rhabdomyolysis in a dengue fever patient: a case report and review of literature. *Rev Inst Med Trop Sao Paulo* 2014;56:85-8.
100. Wiersinga WJ, Scheepstra CG, Kasanardjo JS, et al. Dengue fever-induced hemolytic uremic syndrome. *Clin Infect Dis* 2006;43:800–1
101. Upadhaya BK, Sharma A, Khaira A, et al. Transient IgA nephropathy with acute kidney injury in a patient with dengue fever. *Saudi J Kidney Dis Transpl* 2010;21:521–5.
102. Khalil MA, Tan J, Khalil MA, et al. Predictors of hospital stay and mortality in dengue virus infection-experience from Aga Khan University Hospital Pakistan. *BMC Res Notes*. 2014;7:473.
103. Laoprasopwattana K, Chaimongkol W, Pruekprasert P, et al. Acute respiratory failure and active bleeding are the important fatality predictive factors for severe dengue viral infection. *PLoS One*. 2014;9:e114499.
104. Mallhi TH, Khan AH, Sarriff A, et al. Association of Ward Acquired, On-admission, Progressive and Non-progressive AKI with Death among Dengue Patients: A Hidden Relationship. *Acta Med Port*. 2016;29:157-8.
105. Wills BA, Nguyen MD, Ha TL, et al. Comparison of three fluid solutions for resuscitation in dengue shock syndrome. *N Engl J Med*. 2005;353:877-89.
106. Zhang F, Kramer CV. Corticosteroids for dengue infection. *Cochrane Database Syst Rev*. 2014; 7:CD003488.
107. Watt G, Parola P. Scrub typhus and tropical rickettsioses. *Curr Opin Infect Dis*. 2003;16:429-36.

108. WHO Health Organization Regional Office for South-East Asia. Frequently Asked Questions-Scrub Typhus.  
[http://www.searo.who.int/entity/emerging\\_diseases/CDS\\_faq\\_Scrub\\_Typhus.pdf](http://www.searo.who.int/entity/emerging_diseases/CDS_faq_Scrub_Typhus.pdf)) accessed on June 22, 2016.
109. Balcells ME, Rabagliati R, García P, et al. Endemic scrub typhus-like illness, Chile. *Emerg Infect Dis.* 2011;17:1659-63.
110. Kumar M, Krishnamurthy S, Delhikumar CG, et al. Scrub typhus in children at a tertiary hospital in southern India: clinical profile and complications. *J Infect Public Health.* 2012;5:82-8.
111. Paris DH, Shelite TR, Day NP, et al. Unresolved problems related to scrub typhus: a seriously neglected life-threatening disease. *Am J Trop Med Hyg.* 2013;89:301-7.
112. Vikrant S, Dheer SK, Parashar A, et al. Scrub typhus associated acute kidney injury - a study from a tertiary care hospital from western Himalayan State of India. *Ren Fail.* 2013;35:1338-43.
113. Sinha P, Gupta S, Dawra R, et al. Recent outbreak of scrub typhus in North Western part of India. *Indian J Med Microbiol.* 2014;32:247-50.
114. Loomba V, Mani A, John M, et al. Scrub typhus in Punjab: an acute febrile illness with multisystem involvement. *Trop Doct.* 2014;44:152-5.
115. Sharma N, Biswal M, Kumar A et al. Scrub Typhus in a Tertiary Care Hospital in North India. *Am J Trop Med Hyg.* 2016 Jun 13. pii: 16-0086. [Epub ahead of print]
116. Attur RP, Kuppasamy S, Bairy M, et al. Acute kidney injury in scrub typhus. *Clin Exp Nephrol.* 2013;17:725-9.

117. Kumar V, Kumar V, Yadav AK, et al. Scrub typhus is an under-recognized cause of acute febrile illness with acute kidney injury in India. *PLoS Negl Trop Dis*. 2014;8:e2605.
118. Sun IO, Kim MC, Park JW, et al. Clinical characteristics of acute kidney injury in patients with scrub typhus - RIFLE criteria validation. *J Infect Chemother*. 2014;20:93-6.
119. Lee N, Ip M, Wong B, et al. Risk factors associated with life-threatening rickettsial infections. *Am J Trop Med Hyg*. 2008;78:973-8.
120. Kim DM, Kang DW, Kim JO, et al. Acute renal failure due to acute tubular necrosis caused by direct invasion of *Orientia tsutsugamushi*. *J Clin Microbiol*. 2008;46:1548-50.
121. Tseng BY, Yang HH, Liou JH, et al. Immunohistochemical study of scrub typhus: a report of two cases. *Kaohsiung J Med Sci*. 2008;24:92-8.
122. Raoult D, Woodward T, Dumler JS. The history of epidemic typhus. *Infect Dis Clin North Am*. 2004;18:127-40.
123. Young PC, Hae CC, Lee KH, et al. Tsutsugamushi infection-associated acute rhabdomyolysis and acute renal failure. *Korean J Intern Med*. 2003;18:248-50.
124. Lee S, Kang KP, Kim W, et al. A case of acute renal failure, rhabdomyolysis and disseminated intravascular coagulation associated with scrub typhus. *Clin Nephrol*. 2003;60:59-61.
125. Allen AC, Spitz SA. Comparative Study of the Pathology of Scrub Typhus (Tsutsugamushi Disease) and Other Rickettsial Diseases. *Am J Pathol*. 1945;21: 603-81.
126. Hsu GJ, Young T, Peng MY, et al. Acute renal failure associated with scrub typhus: report of a case. *J Formos Med Assoc*. 1993;92:475-7.

127. Chi WC, Huang JJ, Sung JM, et al. Scrub typhus associated with multiorgan failure: a case report. *Scand J Infect Dis.* 1997;29:634-5.
128. Yen TH, Chang CT, Lin JL, et al. Scrub typhus: a frequently overlooked cause of acute renal failure. *Ren Fail.* 2003;25:397-410.
129. Tomori O. Yellow fever: the recurring plague. *Crit Rev Clin Lab Sci.* 2004;41:391-427.
130. Monath TP, Vasconcelos PF. Yellow fever. *J Clin Virol.* 2015;64:160-73.
131. Wasserman S, Tambyah PA, Lim PL. Yellow fever cases in Asia: primed for an epidemic. *Int J Infect Dis.* 2016;48:98-103.
132. WHO Media centre – Yellow fever fact sheet updated May 2016  
<http://www.who.int/mediacentre/factsheets/fs100/en/> accessed in June 23, 2016.
133. Butler D. Fears rise over yellow fever's next move. *Nature.* 2016;532(7598):155-6.
134. Lima EQ, Nogueira ML. Viral hemorrhagic fever-induced acute kidney injury. *Semin Nephrol.* 2008;28:409-15.
135. Gardner CL, Ryman KD. Yellow fever: a reemerging threat. *Clin Lab Med.* 2010;30:237-60.
136. Barnett ED. Yellow fever: epidemiology and prevention. *Clin Infect Dis* 2007;44:850-6.
137. Vasconcelos PF. [Yellow Fever]. *Rev Soc Bras Med Trop.* 2003;36:275-93.
138. Sérié C, Lindrec A, Poirier A, et al. [Studies on yellow fever in Ethiopia. I. Introduction-clinical symptoms of yellow fever]. *Bull World Health Organ.* 1968;38:835-41.
139. Monath TP, Brinker KR, Chandler FW, et al. Pathophysiologic correlations in a rhesus monkey model of yellow fever with special observations on the acute necrosis of B cell areas of lymphoid tissues. *Am J Trop Med Hyg.* 1981;30:431-43.

140. De Brito T, Siqueira SA, Santos RT, et al. Human fatal yellow fever. Immunohistochemical detection of viral antigens in the liver, kidney and heart. *Pathol Res Pract.* 1992;188:177-181.
141. Vasconcelos PF, Luna EJ, Galler R, et al. Brazilian Yellow Fever Vaccine Evaluation Group. Serious adverse events associated with yellow fever 17DD vaccine in Brazil: a report of two cases. *Lancet.* 2001;358:91-97.
142. Monath TP, Barrett AD. Pathogenesis and pathophysiology of yellow fever. *Adv Virus Res.* 2003;60:343-95.
143. Gershy-Damet GM. [Ultrastructural changes caused by yellow fever virus at the level of the kidneys in newborn mice]. *Arch Inst Pasteur Tunis.* 1984; 61:97-106.
144. Li G, Duan T, Wu X, et al. Yellow fever virus infection in Syrian golden hamsters: relationship between cytokine expression and pathologic changes. *Int J Clin Exp Pathol.* 2008; 1:169-79.
145. Engelmann F, Josset L, Girke T, et al. Pathophysiologic and transcriptomic analyses of viscerotropic yellow fever in a rhesus macaque model. *PLoS Negl Trop Dis.* 2014;8:e3295.
146. <http://www.who.int/mediacentre/factsheets/fs337/en/> accessed on 28/10/2015.
147. Kasturiratne A, Wickremasinghe AR, de Silva N, et al. The global burden of snakebite: a literature analysis and modelling based on regional estimates of envenoming and deaths. *PLoS Med.* 2008;5(11):e218.
148. Harrison RA, Hargreaves A, Wagstaff SC, et al. Snake envenoming: a disease of poverty. *PLoS Negl Trop Dis.* 2009;3(12):e569.
149. de Silva HJ, Kasturiratne A, Pathmeswaran A, et al. Snakebite: the true disease burden has yet to be determined. *Ceylon Med J.* 2013;58:93-5.

150. Warrell DA. Snake bite. *Lancet*. 2010;375:77-88.
151. David S, Matathia S, Christopher S. Mortality predictors of snake bite envenomation in southern India--a ten-year retrospective audit of 533 patients. *J Med Toxicol*. 2012;8:118-23.
152. Ribeiro LA, Albuquerque MJ, de Campos VA, et al. [Deaths caused by venomous snakes in the State of São Paulo: evaluation of 43 cases from 1988 to 1993]. *Rev Assoc Med Bras*. 1998;44:312-8.
153. Kanjanabuch T, Sitprija V. Snakebite nephrotoxicity in Asia. *Semin Nephrol*. 2008;28:363-72.
154. Pinho FM, Yu L, Burdmann EA. Snakebite-induced acute kidney injury in Latin America. *Semin Nephrol*. 2008;28:354-62.
155. Alirol E, Sharma SK, Bawaskar HS, et al. Snake bite in South Asia: a review. *PLoS Negl Trop Dis*. 2010;4:e603.
156. Gutiérrez JM. [Snakebite poisoning in Latin America and the Caribbean: An integral view from a regional perspective]. *Bol. Mal. Salud Amb*. 2011;51:1-16.
157. Rodrigues Sgrignolli L, Florido Mendes GE, Carlos CP, et al. Acute kidney injury caused by bothrops snake venom. *Nephron Clin Pract*. 2011;119:c131-6.
158. Dolab JA, de Roodt AR, de Titto EH, et al. Epidemiology of snakebite and use of antivenom in Argentina. *Trans R Soc Trop Med Hyg*. 2014;108:269-76.
159. Cardi BA, Andrade HF Jr, et al. Differential biodistribution of native and 2 kGy <sup>60</sup>Co irradiated crotoxin in tissues of CBA/J mice. *Nat Toxins*. 1998;6:19-25
160. Boni-Mitake M, Costa H, Vassilieff VS, Rogero JR. Distribution of (125)I-labeled crotoxin in mice tissues. *Toxicon*. 2006 Oct;48(5):550-5.

161. Mello SM, Linardi A, Rennó AL, Tarsitano CA, Pereira EM, Hyslop S. Renal kinetics of *Bothrops alternatus* (Urutu) snake venom in rats. *Toxicon*. 2010 Feb-Mar;55(2-3):470-80.
162. Pinho FM, Zanetta DM, Burdmann EA. Acute renal failure after *Crotalus durissus* snakebite: a prospective survey on 100 patients. *Kidney Int*. 2005;67:659-67.
163. Kularatne SA. Epidemiology and clinical picture of the Russell's viper (*Daboia russelii russelii*) bite in Anuradhapura, Sri Lanka: a prospective study of 336 patients. *Southeast Asian J Trop Med Public Health*. 2003;34:855-62.
164. Krishnamurthy S, Gunasekaran K, Mahadevan S, et al. Russells Viper Envenomation-associated Acute Kidney Injury in Children in Southern India. *Indian Pediatr*. 2015;52:583-6.
165. Kularatne SA, Silva A, Weerakoon K, et al. Revisiting Russell's viper (*Daboia russelii*) bite in Sri Lanka: is abdominal pain an early feature of systemic envenoming? *PLoS One* 2014;9:e90198.
166. Azevedo-Marques MM, Hering SE, Cupo P. Evidence that *Crotalus durissus terrificus* (South American rattlesnake) envenomation in humans causes myolysis rather than hemolysis. *Toxicon*. 1987;25:1163-8.
167. Silveira PV, Nishioka S de A. South American rattlesnake bite in a Brazilian teaching hospital. Clinical and epidemiological study of 87 cases, with analysis of factors predictive of renal failure. *Trans R Soc Trop Med Hyg*. 1992;86:562-4.
168. Bucarechi F, Herrera SR, Hyslop S, Baracat EC, Vieira RJ. Snakebites by *Crotalus durissus* ssp in children in Campinas, São Paulo, Brazil. *Rev Inst Med Trop Sao Paulo*. 2002;44:133-8.
169. Cupo P, Marques MM, Hering SE. [Crotalid bites in children: clinical, laboratory, epidemiologic aspects and treatment approach]. *Rev Soc Bras Med Trop*. 1991;24:87-96.

170. Albuquerque PL, Silva Junior GB, Jacinto CN, et al. Epidemiological profile of snakebite accidents in a metropolitan area of northeast Brazil. *Rev Inst Med Trop Sao Paulo*. 2013;55:347-51.
171. Niraj M, Jayaweera JL, Kumara IW, et al. Acute myocardial infarction following a Russell's viper bite: a case report. *Int Arch Med*. 2013;6:7.
172. Isbister GK, Maduwage K, Scorgie FE, et al. Venom Concentrations and Clotting Factor Levels in a Prospective Cohort of Russell's Viper Bites with Coagulopathy. *PLoS Negl Trop Dis*. 2015;9:e0003968.
173. Silva A, Maduwage K, Sedgwick M, et al. Neurotoxicity in Russell's viper (*Daboia russelii*) envenoming in Sri Lanka: a clinical and neurophysiological study. *Clin Toxicol (Phila)*. 2016;54:411-9.
174. Rajagopala S, Thabah MM, Ariga KK, et al. Acute hypopituitarism complicating Russell's viper envenomation: case series and systematic review. *QJM*. 2015;108:719-28.
175. Saikia D, Majumdar S, Mukherjee AK. Mechanism of in vivo anticoagulant and haemolytic activity by a neutral phospholipase A(2) purified from *Daboia russelii russelii* venom: correlation with clinical manifestations in Russell's Viper envenomed patients. *Toxicon*. 2013;76:291-300.
176. Senise LV, Yamashita KM, Santoro ML. Bothrops jararaca envenomation: Pathogenesis of hemostatic disturbances and intravascular hemolysis. *Exp Biol Med (Maywood)*. 2015;240:1528-36.
177. Pinho FM, Burdmann EA. Fatal cerebral hemorrhage and acute renal failure after young *Bothrops jararacussu* snake bite. *Ren Fail*. 2001;23:269-77.

178. Sinha R, Nandi M, Tullus K, et al. Ten-year follow-up of children after acute renal failure from a developing country. *Nephrol Dial Transplant*. 2009;24:829-33.
179. Herath HM, Wazil AW, Abeysekera DT et al. Chronic kidney disease in snake envenomed patients with acute kidney injury in Sri Lanka: a descriptive study. *Postgrad Med J*. 2012;88:138-42.
180. Waikhom R, Sircar D, Patil K, et al. Long-term renal outcome of snake bite and acute kidney injury: a single-center experience. *Ren Fail*. 2012;34:271-4
181. Silva A, Samarasinghe R, Pilapitiya S, et al. Viper bites complicate chronic agrochemical nephropathy in rural Sri Lanka. *J Venom Anim Toxins Incl Trop Dis*. 2014;20:33.
182. Soe S, Win MM, Htwe TT, et al. Renal histopathology following Russell's viper (*Vipera russelli*) bite. *Southeast Asian J Trop Med Public Health*. 1993;24:193-7.
183. Burdmann EA, Woronik V, Prado EB, et al. Snakebite-induced acute renal failure: an experimental model. *Am J Trop Med Hyg*. 1993;48:82-8.
184. Milani Júnior R, Jorge MT, de Campos FP, et al. Snake bites by the jararacuçu (*Bothrops jararacussu*): clinicopathological studies of 29 proven cases in São Paulo State, Brazil. *QJM*. 1997;90:323-34.
185. Mello CP, Morais IC, Menezes RR, et al. *Bothropoides insularis* venom cytotoxicity in renal tubular epithelia cells. *Toxicon*. 2014;88:107-14.
186. Amaral CF, Da Silva OA, Goody P, et al. Renal cortical necrosis following *Bothrops jararaca* and *B. jararacussu* snake bite. *Toxicon*. 1985;23:877-85.
187. Gundappa RK, Sud K, Kohli HS, et al. Snakebite induced acute interstitial nephritis: report of a rare entity. *Ren Fail*. 2002;24:369-7.

188. Golay V, Roychowdhary A, Pandey R, et al. Acute interstitial nephritis in patients with viperine snake bite: single center experience of a rare presentation. *Saudi J Kidney Dis Transpl.* 2012;23:1262-7.
189. Merchant MR, Khanna UB, Almeida AF, et al. Clinicopathological study of acute renal failure following viperine snake bite. *J Assoc Physicians India.* 1989;37:430-3.
190. de Castro I, Burdmann EA, Seguro AC, et al. Bothrops venom induces direct renal tubular injury: role for lipid peroxidation and prevention by antivenom. *Toxicon.* 2004;43:833-9.
191. Martins AM, Nobre AC, Almeida AC, et al. Thalidomide and pentoxifylline block the renal effects of supernatants of macrophages activated with *Crotalus durissus cascavella* venom. *Braz J Med Biol Res.* 2004;37:1525-30.
192. Nascimento JM, Franchi GC Jr, Nowill AE et al. Cytoskeletal rearrangement and cell death induced by *Bothrops alternatus* snake venom in cultured Madin-Darby canine kidney cells. *Biochem Cell Biol.* 2007;85:591-605.
193. Hernández Cruz A, Barbosa Navarro L, Mendonça RZ, et al. Inflammatory mediators release in urine from mice injected with *Crotalus durissus terrificus* venom. *Mediators Inflamm.* 2011;2011:103193.
194. Frezzatti R, Silveira PF. Allopurinol reduces the lethality associated with acute renal failure induced by *Crotalus durissus terrificus* snake venom: comparison with probenecid. *PLoS Negl Trop Dis.* 2011;5:e1312.
195. Zornetta I, Caccin P, Fernandez J, et al. Envenomations by *Bothrops* and *Crotalus* snakes induce the release of mitochondrial alarmins. *PLoS Negl Trop Dis.* 2012;6:e1526.

196. Collares-Buzato CB, da Cruz-Höfling MA. Disarray of glomerular and tubular cell adhesion molecules in the course of experimental Bothrops moojeni envenomation. *Toxicon*. 2014;78:41-6.
197. Morais IC, Pereira GJ, Orzáez M, et al. L-Aminoacid Oxidase from Bothrops leucurus Venom Induces Nephrotoxicity via Apoptosis and Necrosis. *PLoS One*. 2015;10:e0132569.
198. de Sousa FC, Jorge AR, de Menezes RR, et al. Bothrops erythromelas venom induces apoptosis on renal tubular epithelial cells. *Toxicon*. 2016;118:82-5.
199. Willinger CC, Thamaree S, Schramek H, et al. In vitro nephrotoxicity of Russell's viper venom. *Kidney Int*. 1995;47:518-28.
200. Mandal S, Bhattacharyya D. Ability of a small, basic protein isolated from Russell's viper venom (*Daboia russelli russelli*) to induce renal tubular necrosis in mice. *Toxicon*. 2007;50:236-50.
201. Mitmoonpitak C, Chulasugandha P, Khoo O, et al. Effects of phospholipase A2 and metalloprotease fractions of Russell's viper venom on cytokines and renal hemodynamics in dogs. *Toxicon*. 2013;61:47-53.
202. Suntravat M, Yusuksawad M, Sereemasun A, et al. Effect of purified Russell's viper venom-factor X activator (RVV-X) on renal hemodynamics, renal functions, and coagulopathy in rats. *Toxicon*. 2011;58:230-8.
203. Sunitha K, Hemshekhar M, Thushara RM, et al. Inflammation and oxidative stress in viper bite: an insight within and beyond. *Toxicon*. 2015;98:89-97.
204. Gutiérrez JM, Burnouf T, Harrison RA, et al. A multicomponent strategy to improve the availability of antivenom for treating snakebite envenoming. *Bull World Health Organ*. 2014;92:526-32.

205. Otero R, Gutiérrez J, Beatriz Mesa M, et al. Complications of Bothrops, Porthidium, and Bothriechis snakebites in Colombia. A clinical and epidemiological study of 39 cases attended in a university hospital. *Toxicon* 2002;40:1107-14.
206. Albuquerque PL, Silva GB Jr, Jacinto CN, et al. Acute kidney injury after snakebite accident treated in a Brazilian tertiary care centre. *Nephrology (Carlton)*. 2014;19:764-70.
207. Hung DZ, Yu YJ, Hsu CL et al. Antivenom treatment and renal dysfunction in Russell's viper snakebite in Taiwan: a case series. *Trans R Soc Trop Med Hyg*. 2006;100:489-94.
208. Gutiérrez JM, León G, Lomonte B, et al. Antivenoms for snakebite envenomings. *Inflamm Allergy Drug Targets*. 2011;10:369-80.
209. Martines MS, Mendes MM, Shimizu MH, et al. Effects of Schizolobium parahyba extract on experimental Bothrops venom-induced acute kidney injury. *PLoS One*. 2014;9:e86828.
210. 189. Sharma M, Das D, Iyer JK, et al. Unveiling the complexities of Daboia russelii venom, a medically important snake of India, by tandem mass spectrometry. *Toxicon*. 2015;107(Pt B):266-81.
211. Phillips RE, Theakston RD, Warrell DA, et al. Paralysis, rhabdomyolysis and haemolysis caused by bites of Russell's viper (*Vipera russelli pulchella*) in Sri Lanka: failure of Indian (Haffkine) antivenom. *Q J Med*. 1988;68:691-715.
212. Maduwage K, Silva A, O'Leary MA, et al. Efficacy of Indian polyvalent snake antivenoms against Sri Lankan snake venoms: lethality studies or clinically focussed in vitro studies. *Sci Rep*. 2016;6:26778.
213. Abdulkader RC, Barbaro KC, Barros EJ, et al. Nephrotoxicity of insect and spider venoms in Latin America. *Semin Nephrol*. 2008;28:373-82.

214. Ferreira RS Jr, Almeida RA, Barraviera SR, et al. Historical perspective and human consequences of Africanized bee stings in the Americas. *J Toxicol Environ Health B Crit Rev.* 2012;15:97-108.
215. Rajendiran C, Puvanalingam A, Thangam D, et al. Stroke after multiple bee sting. *J Assoc Physicians India.* 2012;60:122-4.
216. Bilir O, Ersunan G, Kalkan A, et al. A different reason for cerebrovascular disease. *Am J Emerg Med.* 2013;31:891.e5-6.
217. Puttegowda B, Chikkabasavaiah N, Basavappa R, et al. Acute myocardial infarction following honeybee sting. *BMJ Case Rep.* 2014;2014. pii: bcr2014203832.
218. Puvanalingam A, Karpagam P, Sundar C, Venkatesan S, Ragunathanan. Myocardial infarction following bee sting. *J Assoc Physicians India.* 2014;62:738-40.
219. França FO, Benvenuti LA, Fan HW, et al. Severe and fatal mass attacks by 'killer' bees (Africanized honey bees--*Apis mellifera scutellata*) in Brazil: clinicopathological studies with measurement of serum venom concentrations. *Q J Med.* 1994;87:269-82.
220. Deshmukh LS, Borse BT. Acute renal failure following multiple stings by honeybees. *Indian Pediatr.* 1996;33:781-3.
221. Vikrant S, Patial RK. Acute renal failure following multiple honeybee stings. *Indian J Med Sci.* 2006;60:202-4.
222. Deshpande PR, Farooq AK, Bairy M, et al. Acute Renal Failure and/or Rhabdomyolysis due to Multiple Bee Stings: A Retrospective Study. *N Am J Med Sci.* 2013;5:235-9.
223. Mejia G, Arbelaez M, Henao JE, et al. Acute renal failure due to multiple stings by Africanized bees. *Ann Intern Med.* 1986;104:210-1.

224. Hommel D, Bollandard F, Hulin A. Multiple African honeybee stings and acute renal failure. *Nephron*. 1998;78:235-6.
225. Bresolin NL, Carvalho LC, Goes EC, et al. Acute renal failure following massive attack by Africanized bee stings. *Pediatr Nephrol*. 2002;17:625-7.
226. Daher Ede F, da Silva Júnior GB, Bezerra GP, et al. Acute renal failure after massive honeybee stings. *Rev Inst Med Trop São Paulo*. 2003;45:45-50.
227. Gabriel DP, Rodrigues AG Jr, Barsante RC, et al. Severe acute renal failure after massive attack of Africanized bees. *Nephrol Dial Transplant*. 2004;19:2680.
228. Daher Ede F, Oliveira RA, Silva LS, et al. [Acute renal failure following bee stings: case reports]. *Rev Soc Bras Med Trop*. 2009;42:209-12.
229. Mejía Vélez G. [Acute renal failure due to multiple stings by Africanized bees. Report on 43 cases]. *Nefrologia*. 2010;30: 31-8.
230. Bridi RA, Balbi AL, Neves PM, et al. Acute kidney injury after massive attack of Africanised bees. *BMJ Case Rep*. 2014;2014.pii:bcr2013201381.
231. Díaz-Sánchez CL, Lifshitz-Guinzberg A, Ignacio-Ibarra G, et al. Survival after massive (>2000) Africanized honeybee stings. *Arch Intern Med*. 1998;158:925-7
232. Grisotto LS, Mendes GE, Castro I, et al. Mechanisms of bee venom-induced acute renal failure. *Toxicon*. 2006;48:44-54.
233. Prado M, Solano-Trejos G, Lomonte B. Acute physiopathological effects of honeybee (*Apis mellifera*) envenoming by subcutaneous route in a mouse model. *Toxicon*. 2010;56:1007-17.

234. Funayama JC, Pucca MB, Roncolato EC, et al. Production of human antibody fragments binding to melittin and phospholipase A2 in Africanised bee venom: minimising venom toxicity. *Basic Clin Pharmacol Toxicol.* 2012;110:290-7
235. Beccari M, Castiglione A, Cavaliere G, et al. Unusual case of anuria due to African bee stings. *Int J Artif Organs.* 1992;15:281-3.
236. Santos KS, Stephano MA, Marcelino JR, et al. Production of the first effective hyperimmune equine serum antivenom against Africanized bees. *PLoS One.* 2013;8:e79971.
237. Zhang L, Kang Y, Fu P, et al. Myoglobin clearance by continuous venous-venous haemofiltration in rhabdomyolysis with acute kidney injury: a case series. *Injury.* 2012;43:619-23.
238. Sánchez MN, Mignone Chagas MA, Casertano SA, et al. [Accidents with caterpillar *Lonomia obliqua* (Walker, 1855). An emerging problem]. *Medicina (B Aires).* 2015;75:328-33.
239. Arocha-Piñango CL, Larysse M. Fibrinolysis produced by contact with a caterpillar. *Lancet.* 1969;1:810-812.
240. Pinto AF, Berger M, Reck J Jr, et al. *Lonomia obliqua* venom: In vivo effects and molecular aspects associated with the hemorrhagic syndrome. *Toxicon.* 2010;56:1103-12.
241. Arocha-Piñango CL, Marval E, Guerrero B. *Lonomia* genus caterpillar toxins: biochemical aspects. *Biochimie.* 2000;82(9-10):937-42.
242. Duarte AC, Caovilla J., Lorini JD, et al. Insuficiencia renal aguda por accidentes com lagartas. *J Bras Nefrol.* 1990;4:18-7.
243. Burdmann EA, Antunes L, Saldanha LB, et al. Severe acute renal failure induced by the venom of *Lonomia* caterpillars. *Clin Nephrol.* 1996;46:337-9.

244. Fan HW, Cardoso JL, Olmos RD, et al. Hemorrhagic syndrome and acute renal failure in a pregnant woman after contact with *Lonomia* caterpillars: a case report. *Rev Inst Med Trop Sao Paulo*. 1998;40:119-20.
245. Gamborgi GP, Metcalf EB, Barros EJ. Acute renal failure provoked by toxin from caterpillars of the species *Lonomia obliqua*. *Toxicon*. 2006;47:68-74.
246. Da Silva WD, Campos CM, Gonçalves LR, et al. Development of an antivenom against toxins of *Lonomia obliqua* caterpillars. *Toxicon*. 1996;34:1045-9.
247. Seibert CS, Oliveira MR, Gonçalves LR, et al. Intravascular hemolysis induced by *Lonomia obliqua* caterpillar bristle extract: an experimental model of envenomation in rats. *Toxicon*. 2004;44:793-9.
248. Da Silva GH, Panunto PC, Hyslop S, et al. Immunochemical detection of *Lonomia obliqua* caterpillar venom in rats. *Microsc Res Tech*. 2004;65:276-81.
249. Berger M, Santi L, Beys-da-Silva WO, et al. Mechanisms of acute kidney injury induced by experimental *Lonomia obliqua* envenomation. *Arch Toxicol*. 2015;89:459-83.
250. Schmitberger PA, Fernandes TC, Santos RC, et al. Probable chronic renal failure caused by *Lonomia* caterpillar envenomation. *J Venom Anim Toxins Incl Trop Dis*. 2013;19:14.
251. Gremski LH, Trevisan-Silva D, Ferrer VP, et al. Recent advances in the understanding of brown spider venoms: From the biology of spiders to the molecular mechanisms of toxins. *Toxicon*. 2014;83:91-120.
252. Isbister GK, Fan HW. Spider bite. *Lancet*. 2011; 378:2039-47.
253. de Souza AL, Malaque CM, Sztajnbok J, et al. *Loxosceles* venom-induced cytokine activation, hemolysis, and acute kidney injury. *Toxicon*. 2008;51:151-6.

254. Malaque CM, Santoro ML, Cardoso JL, et al. Clinical picture and laboratorial evaluation in human loxoscelism. *Toxicon*. 2011;58:664-71.
255. Golay V, Desai A, Hossain A, et al. Acute kidney injury with pigment nephropathy following spider bite: a rarely reported entity in India. *Ren Fail*. 2013;35:538-40.
256. Nag A, Datta J, Das A, et al. Acute kidney injury and dermonecrosis after *Loxosceles reclusa* envenomation. *Indian J Nephrol*. 2014;24:246-8.
257. Sezerino UM, Zannin M, Coelho LK, et al. A clinical and epidemiological study of *Loxosceles* spider envenoming in Santa Catarina, Brazil. *Trans R Soc Trop Med Hyg*. 1998;92:546-8.
258. Schenone H, Saavedra T, Rojas A, et al. Loxoscelismo en Chile. Estudios epidemiológicos, clínicos y experimentales. *Rev Inst Med Trop Sao Paulo*. 1989;31:403-15.
259. Alcides ZF, Jorge GC, Guillermo CG. Desenlace fatal por loxoscelismo cutáneo visceral: Report of one case. *Rev. méd. Chile*. 2005;133:219-23.
260. Sanabria H, Zavaleta A. Panorama epidemiológico del loxoscelismo en el Perú. *Rev. Perú. med. exp. salud pública* [online]. 1997;14:33-41.
261. Luciano MN, da Silva PH, Chaim OM, et al. Experimental evidence for a direct cytotoxicity of *Loxosceles intermedia* (brown spider) venom in renal tissue. *J Histochem Cytochem*. 2004;52:455-67.
262. Chaim OM, Sade YB, da Silveira RB, et al. Brown spider dermonecrotic toxin directly induces nephrotoxicity. *Toxicol Appl Pharmacol*. 2006;211:64-77.
263. Lucato RV Jr, Abdulkader RC, Barbaro KC, et al. *Loxosceles gaucho* venom-induced acute kidney injury-in vivo and in vitro studies. *PLoS Negl Trop Dis*. 2011;5:e1182.

264. Anwar S, Torosyan R, Ginsberg C, et al. Clinicopathological course of acute kidney injury following brown recluse (*Loxoscles reclusa*) envenomation. *Clin Kidney J*. 2013;6:609-12.
265. Pauli I, Puka J, Gubert IC, et al. The efficacy of antivenom in loxoscelism treatment. *Toxicon*. 2006;48:123-37.
266. Said A, Hmiel P, Goldsmith M, et al. Successful use of plasma exchange for profound hemolysis in a child with loxoscelism. *Pediatrics*. 2014;134:e1464-7.
267. Chippaux JP, Goyffon M. Epidemiology of scorpionism: a global appraisal. *Acta Trop*. 2008;107:71-9.
268. Isbister GK, Bawaskar HS. Scorpion envenomation. *N Engl J Med*. 2014;371:457-63.
269. Viswanathan S, Prabhu C. Scorpion sting nephropathy. *NDT Plus*. 2011;4:376-82.
270. Cupo P. Clinical update on scorpion envenoming. *Rev Soc Bras Med Trop*. 2015;48:642-9.
271. Torrez PP, Quiroga MM, Abati PA, et al. Acute cerebellar dysfunction with neuromuscular manifestations after scorpionism presumably caused by *Tityus obscurus* in Santarém, Pará/Brazil. *Toxicon*. 2015;96:68-73.
272. Angsanakul J, Sitprija V. Scorpion venoms, kidney and potassium. *Toxicon*. 2013;73:81-7.
273. Bucarechi F, Fernandes LC, Fernandes CB, et al. Clinical consequences of *Tityus bahiensis* and *Tityus serrulatus* scorpion stings in the region of Campinas, southeastern Brazil. *Toxicon*. 2014;89:17-25.

274. Revelo MP, Bambirra EA, Ferreira AP, Diniz CR, Chávez-Olórtegui C. Body distribution of *Tityus serrulatus* scorpion venom in mice and effects of scorpion antivenom. *Toxicon*. 1996;34:1119-25.
275. Rajasekhar D, Mohan A. Clinical and echocardiographic findings in patients with myocardial toxicity due to scorpion sting. *Natl Med J India*. 2004;17:307-9.
276. Ranaweera GG, Bavanthan V, Nazar AL, Lokuhetty MD. Acute renal insufficiency after scorpion sting. *Ceylon Med J*. 2015;60:31-2.
277. Saidani C, Hammoudi-Triki D, Laraba-Djebari F, Taub M. In vitro studies with renal proximal tubule cells show direct cytotoxicity of *Androctonus australis hector* scorpion venom triggered by oxidative stress, caspase activation and apoptosis. *Toxicon*. 2016;120:29-37.
278. Silva NA, Albuquerque CM, Marinho AD, et al. Effects of *Tityus stigmurus* (Thorell 1876) (Scorpiones: Buthidae) venom in isolated perfused rat kidneys. *An Acad Bras Cienc*. 2016;88 Suppl 1:665-75.
279. Heidarpour M, Ennaifer E, Ahari H, et al. Histopathological changes induced by *Hemiscorpius lepturus* scorpion venom in mice. *Toxicon*. 2012;59:373-8.
280. Gutiérrez-Mendoza I, Serna-Vela FJ, Góngora-Ortega J, Pérez-Guzmán C, Martínez-Saldaña MC, Loza IL. [Scorpion sting and its relationship with child chronic kidney disease]. *Salud Publica Mex*. 2011;53:106-7.

## Figure legends

Figure 1. Hemorrhagic blister and tissue necrosis at the bite site 48 hours after Bothrops snakebite (Courtesy Dr. Carlos A. C. Mendes, Sao Jose do Rio Preto Medical School, Brazil).

Figure 2. A. Contrast enhanced CT scan of a patient with persistent anuria after bite by a Russell's viper shown acute cortical necrosis characterized by nonenhancing renal cortex bordered on the outside by capsular rim and inside by the enhancing medulla. B. Histological examination of renal biopsy shown necrosis of all elements of renal parenchyma (H&E x 440)

Figure 3. *Lonomia obliqua* caterpillars, with the body covered by bristles, which inject the venom when the victim touches the caterpillar. Note the perfect mimetics of the animal (Courtesy Prof. Elvino J. G. Barros, Rio Grande do Sul Federal University Medical School, Brazil).

Figure 4. Typical local injury three days after *Loxosceles* spider bite (Courtesy Dr. Carlos A. C. Mendes, Sao Jose do Rio Preto Medical School, Brazil).

Table 1. Common differential diagnosis of tropical acute febrile illness associated to AKI.

<b>Clinical picture</b>	<b>Differential diagnosis</b>
Fever + jaundice	Leptospirosis, malaria, dengue, yellow fever, hantavirus, rickettsiosis, acute hepatitis
Biphasic fever + conjunctival suffusion + thrombocytopenia + transaminitis	Leptospirosis
Continuous fever + severe respiratory symptoms leading to ARDS	Hantavirus
Fever + severe myalgia + thrombocytopenia + acalculous cholecystitis	Dengue
Fever + maculopapular rash + 'eschar'	Scrub typhus
Fever + splenomegaly + thrombocytopenia	Malaria
Fever + exposure to unpasteurized milk products	Brucellosis
Fever + diarrhea	Bacterial or viral gastroenteritis

ARDS: acute respiratory distress syndrome.

Table 2. Some characteristics of the main febrile tropical diseases associated to AKI in South Asia and/or Latin America.

	<b>Malaria</b>	<b>Leptospirosis</b>	<b>Dengue</b>	<b>Scrub Typhus</b>	<b>Yellow Fever</b>
<b>Pathogen</b>	<i>Plasmodium</i> protozoans	<i>Leptospira</i> spirochetes	<i>Flavivirus</i> RNA arbovirus	Bacterium <i>Orientia tsutsugamushi</i>	Yellow fever arbovirus
<b>Vector</b>	<i>Anopheles</i> mosquitos	Rodents, dogs, pigs, cattle, horses	<i>Aedes</i> mosquitos	<i>Leptotrombidium</i> mites larva	<i>Haemagogus</i> and <i>Aedes</i> mosquitos
<b>Affected areas</b>	SA and LA	SA and LA	SA and LA	SA	LA
<b>AKI frequency</b>	1 to 60%	40 to 87%	0.9 to 30.7%	20 to 60%	Unknown
<b>Renal histology</b>	ATN, CN, TMA, GN	ATN, AIN, vasculitis	ATN, TMA, GN	ATN, AIN, GN	ATN
<b>Long-term CKD</b>	Described	Described	Unknown	Unknown	Unknown

AKI: acute kidney injury; CKD: chronic kidney disease; SA: South Asia; LA: Latin America; ATN: acute tubular necrosis; CN: cortical necrosis; TMA: thrombotic microangiopathy; GN: glomerulonephritis; AIN: acute interstitial nephritis;

Table 3. Some characteristics of the venomous animals causing accidents associated to AKI in South Asia and/or Latin America

	<b>Russell's Viper</b>	<b>Bothrops</b>	<b>Crotalus</b>	<b>Bees</b>	<b>Lonomia</b>	<b>Loxosceles</b>	<b>Scorpions</b>
<b>Phylum</b>	Cordata	Cordata	Cordata	Arthropoda	Arthropoda	Arthropoda	Arthropoda
<b>Class</b>	Reptilia	Reptilia	Reptilia	Insecta	Insecta	Arachnida	Arachnida
<b>Order</b>	Squamata	Squamata	Squamata	Hymenoptera	Lepidoptera	Arenae	Scorpionida
<b>Suborder</b>	Serpentes	Serpentes	Serpentes	Apocrita	Glossata	Araneomorphae	Scorpiones
<b>Affected areas</b>	SA	LA	LA	SA and LA	LA	SA and LA	SA and LA
<b>Venom effects</b>	Local injury, hemolysis, rhabdomyolysis, coagulopathy, neurotoxicity	Local injury, coagulopathy	Neurotoxicity, rhabdomyolysis	Hemolysis, coagulopathy, rhabdomyolysis, heart, lung, and liver injury, neurotoxicity	Coagulopathy	Local injury, hemolysis, rhabdomyolysis, hypotension and shock	Autonomic overactivity, dysfunction, edema, neuronal excitation
<b>AKI frequency</b>	18.7 to 45 %	1.4 to 38%	10 to 33%	unknown	1.9%	3.3 to 6.4%	unknown
<b>Renal histology</b>	ATN, CN, AIN, GC	ATN, CN, AIN, GC	ATN	ATN	ATN, GC	ATN	ATN, CN, AIN, GC
<b>Long-term CKD</b>	Described	Unknown	Unknown	Unknown	Described	Unknown	Unknown

GD: geographic distribution; SA: South Asia; LA: Latin America; AKI: acute kidney injury; ATN: acute tubular necrosis; CN: cortical necrosis;

AIN: acute interstitial nephritis; GC: glomerular changes; TMA: thrombotic microangiopathy; CKD: chronic kidney disease.